

# Water Screen Assay Card

## Hybridization Probe Assays

This reagent kit is designed to test for *Cryptosporidium*, *E. coli O157*, and *Salmonella*, from two unknown liquid or dry samples. These protocols are to be performed manually and have been simplified to reduce the risk of operator error.

**⚠️ WARNING:** If the unknown sample to be tested is suspected to contain pathogen or toxin, it must be handled by trained personnel and treated as if it were highly poisonous or infectious throughout the sample purification procedure, the assay procedure, and when disposing of all waste materials that have come in contact with the suspected sample.

### Kit Contents

Kit Part No. PATH-ASY-0011

1- 5 mL Reagent Grade Water	2- 5 mL Unknown Sample Bottles	4- 1 mL Syringes with Cannula Tips
1- Sample Swab Pack	1- Instruction Booklet	1- Freeze-dried Reagents in a Foil Bag
1- Transfer Pipettes Pack	1- Loading Instructions Sticker	

Part # RAZR-ASY-0003, RAZR-ASY-0013, and/or RAZR-RED-4000 may be required

## Programming the Run Protocol

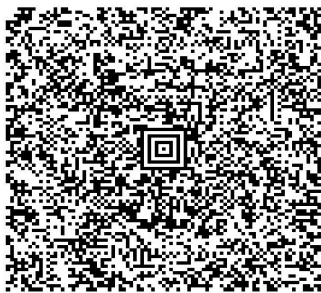
Before the sample is loaded into the pouch, verify that the run protocol for this test is loaded onto the instrument. If it is not and you are using a RAZOR®, use the RAZOR desktop software to create the protocol and load it onto the instrument. Use the data in this card when creating the protocol for this particular test with the Protocol Wizard. For the RAZOR EX, you can load the protocol by scanning the square protocol bar code on the reagent box. If the bar code is damaged, use the generic bar codes that are located below. Times, temperatures, and results analyses are all included in the protocol bar code.

## Loading Protocols for the RAZOR® EX

After the square protocol bar code has been scanned, scan the rectangular bar code on the pouch fitment. If the rectangular bar code is damaged, use the generic one below. **Note: you will only be able to use the generic bar code for one run. In order to re-use it, you have to delete the run.**



WATRSCNA-0000001



Protocol Code: WATRSCNA

### Step 1:

#### Negative Port

Add 0.5 mL reagent grade water

### Step 2:

#### Unknown 1 Port

Add 0.5 mL sample 1

### Step 3:

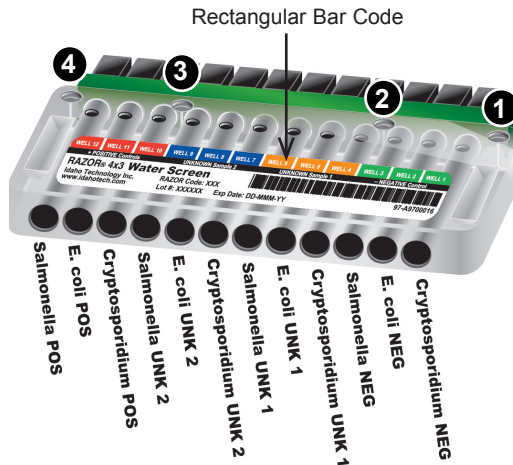
#### Unknown 2 Port

Add 0.5 mL sample 2

### Step 4:

#### Positive Port

Add 0.5 mL reagent grade water



# Protocol Steps for the RAZOR®

Protocol Steps	Data to be Entered																																																																																				
<b>Step 1: Protocol Identifier</b>	Enter: <b>0BH</b> to identify this protocol. This identifying code matches the first three characters on the pouch. This will help you identify the protocol after it is loaded on the instrument.																																																																																				
<b>Step 2: Protocol Parameters</b>	Initial Hold Temperature: <b>93°C</b> Denature Temperature: <b>91°C</b> Anneal Temperature: <b>60°C</b>																																																																																				
	Initial Hold Time: <b>120 s</b> Denature Hold Time: <b>3 s</b> Anneal Hold Time: <b>15 s</b>																																																																																				
	Cycles: <b>55</b>																																																																																				
<b>Step 3: Select Organisms for the Protocol</b>	A list of organisms is listed in the <b>All Organisms</b> column. Select <b>Crypto</b> and click the <b>Add</b> button. Repeat this for <b>E. coli</b> and <b>Salm</b> . All three have just been added to the <b>Organism For This Protocol</b> column. If <b>Crypto</b> , <b>E. coli</b> or <b>Salm</b> is not listed, see the RAZOR Users Manual for instructions on how to add new organisms.																																																																																				
<b>Step 4: Add Environmental Samples</b>	Add two unknown samples by clicking the <b>Add</b> button and entering <b>UNK1</b> . Repeat the steps and enter <b>UNK2</b> .																																																																																				
<b>Step 5: Assign an Organism to Each Sample Position</b>	Assign <b>Crypto</b> to positions 1, 4, 7 and 10. Assign <b>E. coli</b> to positions 2, 5, 8 and 11. Assign <b>Salm</b> to positions 3, 6, 9, and 12.																																																																																				
<b>Step 6: RAZOR Pouch Definition</b>	<p>Fill out the definition as shown in the image.</p> <table border="1"> <thead> <tr> <th colspan="6">Razor Bag Definition</th> </tr> <tr> <th>Pos</th> <th>Organism</th> <th>Target</th> <th>Type</th> <th>Env. Sample</th> <th>Concentration</th> </tr> </thead> <tbody> <tr><td>1</td><td>Crypto</td><td>Target</td><td>NEG</td><td></td><td></td></tr> <tr><td>2</td><td>E. coli</td><td>Target</td><td>NEG</td><td></td><td></td></tr> <tr><td>3</td><td>Salm</td><td>Target</td><td>NEG</td><td></td><td></td></tr> <tr><td>4</td><td>Crypto</td><td>Target</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>5</td><td>E. coli</td><td>Target</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>6</td><td>Salm</td><td>Target</td><td>UNK</td><td>UNK1</td><td></td></tr> <tr><td>7</td><td>Crypto</td><td>Target</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>8</td><td>E. coli</td><td>Target</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>9</td><td>Salm</td><td>Target</td><td>UNK</td><td>UNK2</td><td></td></tr> <tr><td>10</td><td>Crypto</td><td>Target</td><td>POS</td><td></td><td></td></tr> <tr><td>11</td><td>E. coli</td><td>Target</td><td>POS</td><td></td><td></td></tr> <tr><td>12</td><td>Salm</td><td>Target</td><td>POS</td><td></td><td></td></tr> </tbody> </table>	Razor Bag Definition						Pos	Organism	Target	Type	Env. Sample	Concentration	1	Crypto	Target	NEG			2	E. coli	Target	NEG			3	Salm	Target	NEG			4	Crypto	Target	UNK	UNK1		5	E. coli	Target	UNK	UNK1		6	Salm	Target	UNK	UNK1		7	Crypto	Target	UNK	UNK2		8	E. coli	Target	UNK	UNK2		9	Salm	Target	UNK	UNK2		10	Crypto	Target	POS			11	E. coli	Target	POS			12	Salm	Target	POS		
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<b>Step 7: RAZOR Meta Calls</b>	This step displays an image of the conditions required for an unknown to be called positive. You do not need to enter any data in this step. Click <b>Finish</b> to save the protocol.																																																																																				
<b>NOTE:</b>	<p>Before interpreting results on machine's LED screen, remember assays correlate to the channels shown in diagram to right.</p> 